

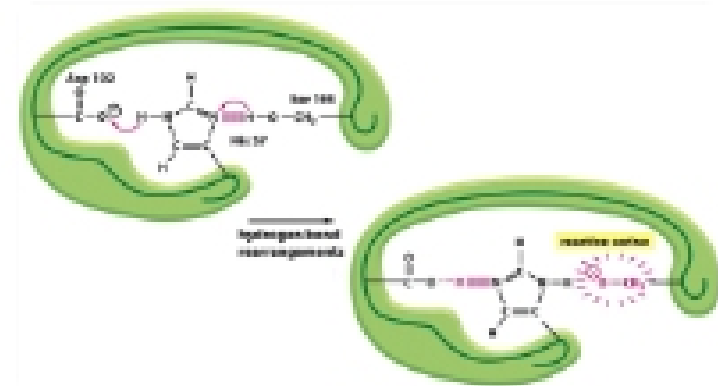
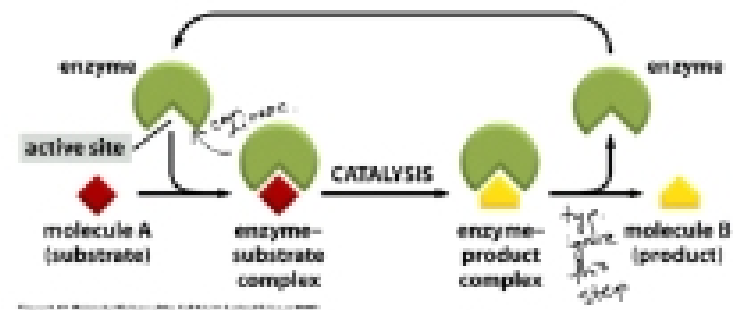
Main Ideas

- Enzymes are catalysts that **lower the activation energy** for a rxn and speed up the rxn rate
- Binding of a protein to a ligand (1st step of catalysis) can be measured by the **binding (equilibrium) constant**
- The Michaelis-Menten (MM) equation relates the rate of an enzyme catalyzed rxn to the **substrate concentration** and two constants, K_m and k_{cat}
- Based on the MM equation, the **best enzymes have a small K_m and a large V_{max} (w/ $K_m = [S]$ at $V_{max}/2$)**
- Ex. of enzymatic amplification thru regulated proteolysis= **blood coagulation cascade**

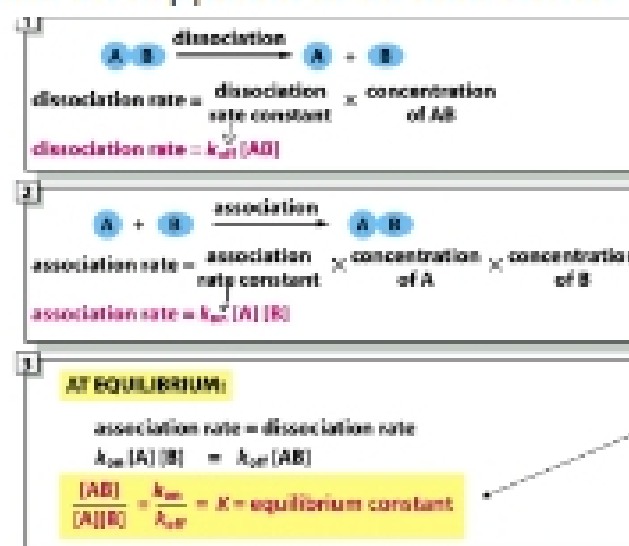
Learning Objectives

1) Understand the basic principles and steps of enzyme catalyzed rxns

- **Enzymes** are catalysts and are everywhere
- Catalyzed rxns are typically connected in series
- Enzymes lower the **activation energy** that blocks chemical rxns --> hugely increase the rate/speed of chemical rxns, but don't affect/change K_{eq} (the state where products and reactants are in equilibrium)
- They are **highly selective**= 1 enzyme: 1 rxn
 - Due to **active site**: pocket only accepts 1 substrate
- **Substrate/ligand**= molecule that binds to the enzyme
 - Binds thru non-covalent interactions and complementary binding surfaces
 - Hypothesis: 1) lock and key= fit each other perfectly
 - 2) induced fit= don't fit perfectly initially, enzyme has to undergo a little change to fit perfectly
- Ex. **serine protease**= attacks peptide bonds on proteins, has a basic microenvi, so due to the interactions between its side chains via H-bonds, a nonreactive group-->reactive group



- Active site: catalytic triad
- First step: **substrate binding**
 - Dynamic process: the substrate binds to other molecules, but their interactions are too weak, so they fall apart. Substrate only stays w/enzyme when it's supposed to be bound to it.
- **Equilibrium constant** measures binding strength
 - at equilibrium the association rate = the dissociation rate
 - $k_1 = k_{on}$
 - $k_2 = k_{cat}$ (catalysis)= $ES \rightarrow E + P$
 - this is the limiting step
 - ES is a transient intermediate



equilibrium association constant

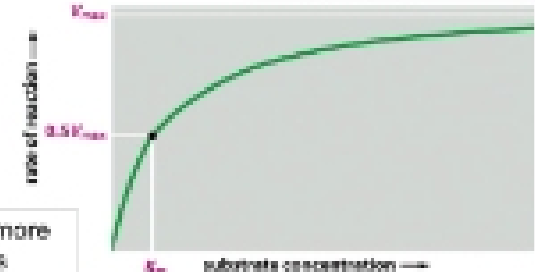
- understand steady state approximation: ES concentration stays the same, so $d[ES]/dt = 0$, helps us get to MM

2) Understand that the MM eqn describes how rxn velocity depends on substrate concentration

Michaelis-Menten Equation

$$v = \frac{d[P]}{dt} = \frac{k_2 [E_1][S]}{K_m + [S]}$$

velocity of reaction



At high [S], adding more substrate does not change the rate much.

At low [S], adding more substrate increases rate significantly.

When $[S] \gg K_m$, then:

$$v = v_{max} = k_2[E_1]$$

- So the velocity of your rxn is the increase in product made over time and this depends on your substrate concentration

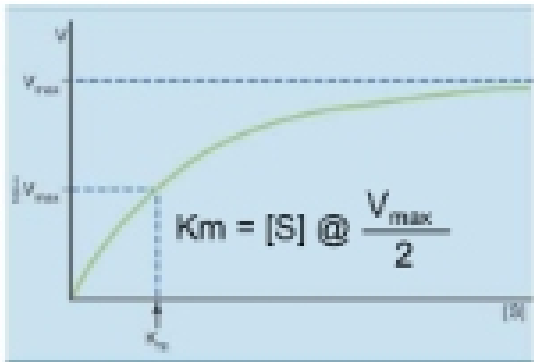
Using the Michaelis constant: $K_m = \frac{k_{-1} + k_2}{k_1}$
(rate of (koff + kcat) / (kon))

- v_{max}
 - once all the enzyme is bound by substrate, the max rxn rate is reached

$$v = \frac{k_2 [E_1][S]}{K_m + [S]} = \frac{v_{max} [S]}{K_m + [S]}$$

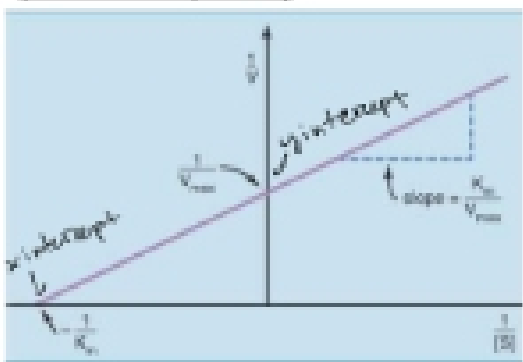
Note: $k_2 = k_{cat}$ (turnover number)

Direct Plot



$$v = \frac{v_{max} [S]}{K_m + [S]}$$

Lineweaver-Burk Plot (double reciprocal)



$$\frac{1}{v} = \frac{1}{v_{max}} + \frac{K_m}{v_{max}} \times \frac{1}{[S]}$$

Direct Plot:

- When your rxn has reached half of its max v, Km = S

If $k_{-1} \gg k_{cat} \Rightarrow K_m \approx K_d$
(dissoc. constant)

Ergo: K_m indicates affinity between substrate and enzyme

- The best enzymes have the smallest K_m (least tendency to dissociate) and largest V_{max} !

3) Be able to interpret a direct plot or a double reciprocal plot for an enzyme that follows MM kinetics

- See above

4) Be familiar w/the basic principle of an enzyme amplification cascade (ex. blood coagulation)

- Enzymes and rate acceleration

- Range: 10^9 - 10^{23} increase in rate
- Factors that can influence rate:
 - Concentration of enzyme and substrate, affinity between E and S, ability of E to stabilize transition state (ES), bond strain, T of rxn and pH of rxn

- Enzyme Cascade: blood coagulation

- Hemostatis: arrest of blood loss from ruptured blood vessels
 - Primary: vascular restriction + platelets adhere to injury site
 - Secondary: **coagulation**= the cascade of rxns that produces a fibrin mesh (clot) that entraps the soft plug from above
- Triggering coagulation
 - Membrane proteins exposed by injury serve as receptors for zymogens and protein co-factors that are either: already present in the blood or released by platelets following their activation

- Both intrinsic and extrinsic cascade using proteolysis to amplify the signal

- Multimembered cascade
- One member gets activated and then activates the next zymogen --> etc.
- One enzyme at the start of the cascade can activate many molecules of a given substrate

- Factors XII, XI, IX, VII, X, and II: zymogens (=precursor) of serine proteases

- Cleavage of 1 zymogen-->active protease that activates another zymogen
- Each zymogen is also reg. by protein cofactors that themselves are also reg. by proteases (+ feedback looping-->more amplification)
- zymogens are cleaved (all these lose their N terminal parts to free up the serine protease part) to become active

